Comunicación corta/ Short report

First characterization of preantral follicles in the ovaries of adult alpaca

Luisa Echevarria¹, Johanna Leiva-Revilla², Roy Andrade¹, Johan Smitz⁴, José Ricardo Figueiredo³

¹Universidad Peruana Cayetanož: UW/ ˈhJX XY A YX]WoU J YhYf]bUf]U mNcchYVb]Už Heredia, Lima, Perú ²Centro para el Desarrollo de la Investigación Científica (CEDIC), Asunción, Paraguay

³I b]j Yfg]XUX 9ghUhU XY 7YUfz ž Laboratorio de Manipulación de Oocitos y Folículos Preantrales. (LAMOFOPA), Fortaleza, CE, Brasil

⁴Follicle Biology Laboratory, Vrije Universiteit Brussel. Laarbeeklaan 101, B-1090 Brussels, Belgium

Cómo referenciar este artículo/ How to reference this article:

Echevarria L, Leiva-Revilla J, Andrade R, Smitz J, Figueiredo J.R. First characterization of preantral follicles in the ovaries of adult alpaca. Mem. Inst. Investig. Cienc. Salud. 2019; 17(2): 102-106

ABSTRACT

It is a fact that while even basic reproductive information on alpacas is unavailable, the normal ovarian reserve of this species in comparison to other species is also unidentified. In this study, the ovarian preantral follicles in healthy adult alpacas were characterized in order to establish a general model to in vitro studies. Ten ovaries were collected from five adult alpacas. The ovarian cortex samples were fixed with paraformaldehyde and histological analysis was done. Normal and degenerated follicles percentages were determined. The normal follicles were measured and classified in primordial, transitional, primary and secondary stages. preantral follicles present in the ovarian cortex of alpacas were primordial and transitional stages; primary (6.10%) and secondary (0.37%) follicles were rarely found. The primary and secondary follicles were larger in diameter when compared with the primordial and transitional follicles. The largest oocyte diameter was recorded in the secondary follicles (P < 0.05). This study serves to establish a biological model for future reproduction studies in Alpacas or as possible biological model for studies of folliculogenesis in humans.

Keywords: Alpaca, follicle, ovary, histology.

Primera caracterización de folículos preantrales de ovario de alpaca adulta

RESUMEN

Es un hecho que, si bien no se dispone de información reproductiva básica sobre las alpacas, la reserva ovárica normal de esta especie en comparación con otras especies tampoco está identificada. En este estudio, se caracterizaron los folículos preantrales ováricos en alpacas adultas sanas. Se recogieron diez ovarios de cinco alpacas adultas. Las muestras de la corteza ovárica se fijaron con paraformaldehído y se realizó un análisis histológico. Se determinaron los porcentajes de folículos normales y degenerados. Los folículos normales se midieron y clasificaron en estadios: primordiales, de transición, primarios y secundarios. La mayoría de los folículos preantrales presentes en la corteza ovárica de las alpacas eran estadios primordiales y de transición; Raras veces se encontraron folículos primarios (6.10%) y secundarios

Fecha de recepción: mayo 2019. Fecha de aceptación: junio 2019

*Autor correspondiente: Dr. Johanna Leiva Revilla. Center for the Development of Scientific Research (CEDIC), Asuncion, Paraguay. Email: johileiva@gmail.com

(0.37%). Los folículos primarios y secundarios tenían un diámetro mayor en comparación con los folículos primordiales y de transición. El mayor diámetro de ovocitos se registró en los folículos secundarios (P <0.05). Este estudio sirve para establecer un modelo biológico para futuros estudios de reproducción en alpacas o como posible modelo biológico para estudios de foliculogénesis en humanos.

Palabras clave: Alpaca; Folículo; Ovario; Histología

INTRODUCTION

Alpaca (Vicugna pacos), is one of the South American camelids of Peru. The main reproductive characteristics of the alpaca include induced ovulation by mating, very high embryo mortality rate and an 11-month gestation period (1). The ovaries are oval in shape, and 1.5 to 2.5 cm long and 1.0 to 1.2 cm high in size (2). The follicular waves, normally present, last between 9 and 22 days. Spontaneous corpus luteum (CL) formation is rare, and only 5% of the alpacas spontaneously ovulate (3).

The preantral follicles represent the main ovarian reserve. Their activation and arrest stages during folliculogenesis are poorly understood (4). Considering the significance of the preantral follicles in assisted reproductive techniques, several studies have been conducted to describe their morphometric aspects in cow, goats, ewe and llamas (5–8). However, to the best of our knowledge, no reports are yet available regarding the ovarian preantral follicles in the alpaca. Establishing the follicular population of the ovary of adult alpacas could help us to see the possibility of using it as a biological model for studies of folliculogenesis in humans.

The aim of this study was to characterize the preantral follicle population in the alpaca ovary, investigating the following endpoints: follicular morphology (normal vs degenerated), proportion of the primordial, transitional, primary and secondary follicles, as well as the diameters of the follicle and oocyte.

MATERIALS AND METHODS

Animals and ovaries

The methodology, code 102007, was approved by the Institutional Ethics Committee for the Use of Animals (CIEA) of the Universidad Peruana Cayetano Heredia. Ten ovaries from five non-pregnant adult alpacas (2 to 3 years old), with a previous partum, were obtained via median laparotomy cranial to mammary gland, using general anesthesia protocol with Xylazine (1mg/Kg IM), Tramadol (1mg/kg IM) and Ketamine 10% (5mg/kg EV). The ovarian cortical samples were cut into fragments of approximately 3 x 3 x 1 mm sized fragments.

Histological processing

Once harvested, the ovarian tissue was fixed in paraformaldehyde solution at 4 °C for 4 h; then it was placed in 70% alcohol for 24 hours (6) and submitted to histological analysis. The ovarian fragments were dehydrated by passing them through a graded series of alcohol, embedded in paraffin wax for 4 hours at 58 °C, and then sectioned serially into 7 μ m thick slices. The samples were stained using Harris hematoxylin and eosin.

Follicle morphology and development

The histological samples were analyzed using LEICA light microscopy (Wetzlar, Germany) at 400x magnification. The histological evaluation and classification of the follicles were done by a single operator. Based on their morphology, the follicles were categorized as normal (follicle containing an intact oocyte and granulosa cells, well organized in layers, without pyknotic nucleus) or degenerated (oocyte with pyknotic nucleus and retracted cytoplasm, or disorganized granulosa cells detached from the basement membrane) (9,10). Every follicle containing a visible oocyte nucleus was examined in each section where it appeared in and matched with the identical follicle on the adjacent sections to prevent double counting. The follicle and oocyte diameters were measured only in the morphologically normal follicles, using the LEICA software coupled with a LEICA light microscope. The follicle diameter was measured from one

edge of the outermost granulosa cell layer to the opposite edge. Oocyte diameter was measured from one edge of the oocyte membrane to the other. Two perpendicular diameters were recorded for each measurement, and the average of both those values was calculated. Based on their developmental stages the morphologically normal preantral follicles were classified as primordial and developing follicles (transitional, primary and secondary), as reported by Silva-Santos et al., (12).

Statistical analysis

The number of follicles at all stages, primordial and developing (transitional, primary, or secondary), were expressed as mean \pm SD. The diameters of the primordial, transitional, primary and secondary follicles, and oocytes were analyzed using the analysis of variance (ANOVA), and Duncan's mean test.

RESULTS

A total of 624 preantral follicles were evaluated. The normal and degenerated follicles are shown in Figure 1. The overall percentage of the normal preantral follicles was 86.70%. The preantral follicle population showed the following distribution: 71.16% primordial, 22.37% transitional, 6.10% primary, and 0.37% secondary. The follicular and oocyte diameters are shown in Table 1. The primordial and transitional follicles showed similar diameters, which progressively and significantly increased, until they became secondary follicles. The largest oocyte diameter was found in the secondary follicles (P < 0.05).

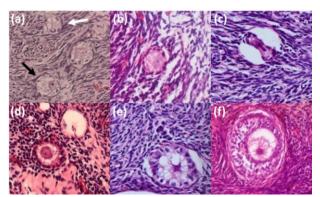


Figure 1: Photomicrographs of the alpaca preantral follicles depicting normal (black arrow) and degenerated (white arrow) primordial follicles (a) normal (b) and degenerated (c) transitional follicles; normal (d) and degenerated (e) primary follicles and normal secondary follicles (f) (magnification 400X).

Table 1: Diameter (μ m) of the preantral follicles (mean \pm SD) and oocytes in the alpaca ovarian cortex samples.

Follicle category	n	Diameter (μm) (mean <u>+</u> SD)	
		Follicle	Oocyte
Primordial	385	40.48 <u>+</u> 5.4 ^a	31.77 <u>+</u> 4.9 ^a
Transitional	121	43.06 <u>+</u> 4.97 ^a	33.08 <u>+</u> 4.1 ^a
Primary	33	49.19 <u>+</u> 8.4 ^b	35.39 <u>+</u> 7.37 ^a
Secondary	2	134.67 <u>+</u> 50 ^c	49.07 <u>+</u> 1.4 ^ь

a,b,c Within a column, values lacking a common superscript differed (p < 0.05).

DISCUSSION

This study is the first to report the histologic evaluation of the preantral follicles of the ovaries of adult alpaca. The occurrence of normal follicles was 86.72%, which concurs with the earlier reports for goats; (91.7%) (13).

The proportions of primordial, transitional, primary and secondary follicles were determined in the present study. Interestingly, the percentages of primordial

(71.23%) and transitional follicles in alpaca were similar to those reported for small ruminants (goat (6) and ewe (7)), while the proportion of the primary follicles (6.05%) corresponded to the findings in bovines (5). Also, Arias et al. (8), in their analysis of ovarian samples of the llama species (Lama glama), found that most preantral follicles were in the primordial state.

The diameter of the alpaca primordial follicles (40.48 μ m) showed similarity to the other species, such as llama (23 – 49.4 μ m; (8)), cow (45.4 μ m; (5) and ewe (33 μ m; (7)). Alpaca's primary follicles (49.19 + 8.4 μ m) were larger than the primordial and transitional ones, but smaller than the llama's primary follicles (62.4 to 75.4 μ m; (8)). Similar results were recorded for the secondary follicles.

Oocyte diameters varied from 31.77 to 49.07 μm and showed similarity among the primordial, transitional and primary follicles; however, they were significantly different from the secondary follicles. Comparable results for oocyte diameters (31.2 and 49.5 μm , for primordial and secondary follicles, respectively) were also described in cow (5); (4). Nevertheless, for both follicular stages, smaller oocyte diameters were observed in alpacas compared to llamas (90 μm) (8).

In conclusion, this is the first study on the preantral follicle population in the adult alpaca ovarian cortex. From the results, it is evident that most of the preantral follicles in the alpaca ovarian cortex were in the primordial or transitional stages and a low rate of follicle degeneration was observed. Regarding the follicle and oocyte diameters, when compared with the literature available on the preantral follicles of llama, the other south American camelid, smaller follicular (primary and secondary follicles) and oocyte (enclosed in primordial and secondary follicles) diameters were found in alpaca, in the current study. This study serves to establish a biological model for future reproduction studies in the Alpacas. Likewise, it serves as a basis for the use of the alpaca ovary as an artificial ovarian model for the human ovary to analyze in vitro folliculogenesis.

FUNDING AND CONFLICT OF INTEREST

This research did not receive any specific funding. The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

JLV would like to thank PRONII-CONACYT (National Research Incentive Program of the National Council for Science and Technology). Paraguay.

BIBLIOGRAPHIC REFERENCES

- Bravo PW, Stabenfeldt GH, Lasley BL, Fowler ME. The effect of ovarian follicle size on pituitary and ovarian responses to copulation in domesticated South American camelids. Biol Reprod. 1991 Oct;45(4):553-9.
- Sumar JB. Reproduction in female South American domestic camelids. J Reprod Fertil Suppl. 1999;54: 169–78.
- Vaughan JL, Tibary A. Reproduction in female South American camelids: A review and clinical observations. Small Rumin Res. 2006 Feb;61(2-3):259-81.
- 4. Fortune J. The early stages of follicular development: activation of primordial follicles and growth of preantral follicles. Anim Reprod Sci. 2003 Oct;78(3-4):135-63.
- van Wezel IL, Rodgers RJ. Morphological characterization of bovine primordial follicles and their

- environment in vivo. Biol Reprod. 1996 Nov;55(5):1003-11.
- Araújo VR, Gomes CM, Deborah M, Modesto G, Báo SN, Viana JR, et al. Effect of Bone Morphogenetic Protein-7 (BMP-7) on in vitro survival of caprine preantral follicles. Pesq Vet Bras. 2010;30(4):305-10.
- 7. Cahill LP, Mauléon P. A study of the population of primordial and small follicles in the sheep. J Reprod Fertil. 1981 Jan;61(1):201–6.
- 8. Arias P, Feder F-H, Porcel R, Cox J. Estudio histológico comparativo del aparato reproductivo de llama hembra (Lama guanicoe glama). Anat Histol Embryol. 1992 Dec;21(4):314–23.
- Amorim CA, Rondina D, Rodrigues APR, Costa SHF, Gonçalves PBD, de Figueiredo JR, et al. Isolated ovine primordial follicles cryopreserved in different concentrations of ethylene

- glycol. Theriogenology. 2003 Sep;60(4):735–42.
- Figueiredo JR, Hulshof SC., Van den Hurk R, Ectors FJ, Fontes RS, Nusgens B, et al. Development of a combined new mechanical and enzymatic method for the isolation of intact preantral follicles from fetal, calf and adult bovine ovaries. Theriogenology. 1993 Oct;40(4):789-99.
- 11. Silva-Santos KC, Santos GMG, Siloto LS, Hertel MF, Andrade ER, Rubin MIB, et al. Estimate of the population of preantral follicles in the ovaries of Bos taurus indicus and Bos taurus taurus cattle. Theriogenology. 2011 Oct;76(6):1051-7.
- 12. Magalhães-Padilha DM, Andrade PM, Sales ET, Araujo VR, Lima IMT, Castro S V, et al. Effect of sequential medium on in vitro culture of goat ovarian cortical tissue. Anim Reprod Sci. 2012 Jun;132(3-4):159-68.